# Automatic design of digital synthetic gene circuits

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## Summary

- Automatic gene circuit design: the problem.
- The Karnaugh map method in biology.
- Comparison with a different design.
- Circuit complexity and performance.
- Conclusion and future work.

## Automatic gene circuit design: previous approaches.

- Given the output, how to derive the corresponding circuit (structure and parameter values?)
- Brute force optimization via evolutionary algorithm (François and Hakim, *PNAS* **101**, 580, 2004)
- Similar implementations: OptCircuit (Dasika and Maranas, *BMC Syst. Bio.* **2**, 24, 2008); Genetdes (Rodrigo *et al., Bioinformatics* **23**, 1857, 2007).

#### **Problems:**

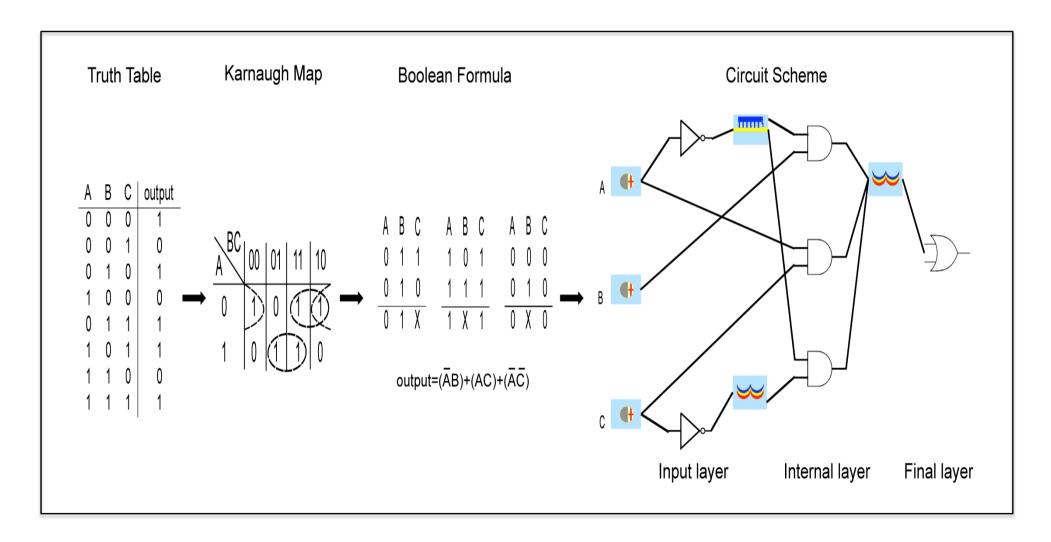
- Transcription units as bio-bricks (instead of parts).
- Limited model (translation as single-step event).
- Double optimization procedure: long computational time.

Looking for a different strategy.

## Digital gene circuits

- Input/Output relation fully described by a truth table.
- The Karnaugh map method converts a truth table into a circuit scheme – no optimization required.
- Boolean gates due to promoter and RBS regulation mechanisms.
- Important application as biosensors.

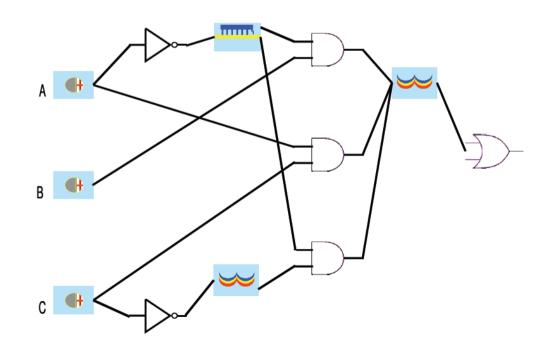
## The Karnaugh map method



Circuit **structure** in three layers – **No optimization** required

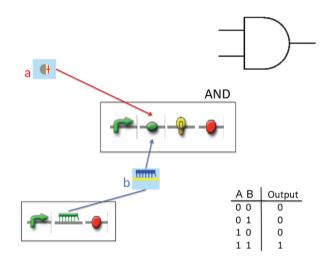
#### Circuit characteristics

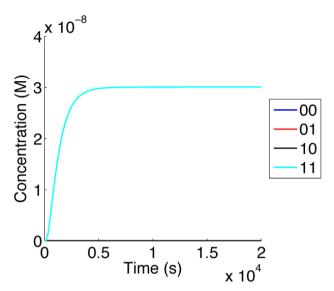
- Activated/Repressed Promoters and RBSs (Bintu et al., Curr. Opin. Genet. Dev. 15, 125, 2005; Isaacs et al., Nat. Biotech. 22, 841, 2004).
- Pools of transcription factors, sRNAs, and chemicals
   (M.A. Marchisio and J. Stelling, "Computational design of synthetic gene circuits with composable parts." Bioinformatics, 24, 1903, 2008).
- A circuit takes up to four inputs (chemicals) and produces a single output (fluorescent protein).



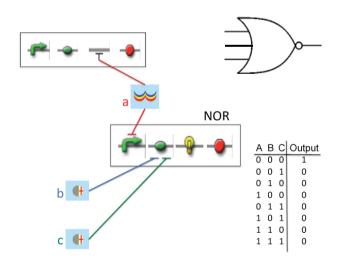
### Gate structure and new designs.

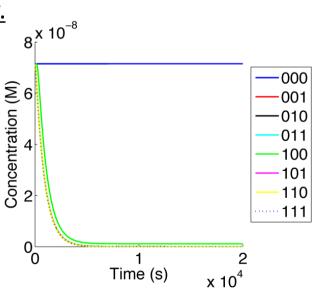
•Riboswitches + sRNA on the RBS.





•Promoters and RBS are controlled simultaneously.





### Comparison with electronics

- Every truth table corresponds to two Boolean formulas: CNF (POS) and DNF (SOP).
- In electronics the minimal circuit is given by the formula with the lowest number of clauses of NOT operations.
- In biology several circuit schemes arise from the same Boolean formula.
- How to define a minimal circuit in biology?

### The complexity score

- Regulatory factors matter more than gene number.
- Only a handful of repressors and activators is currently used.
- Engineering new proteins is more difficult than synthesizing antisense small RNAs.
- Riboswitches simplify the structure of a gate.
- We define as minimal the circuit with the lowest complexity score defined as

$$S = 2^{R-1} + 2^{A-1} + n$$

where: R, repressor number (>= 1); A, activator number (>=1),n antisense sRNA number

- A circuit should avoid to re-use the same kind of transcription factors and prefer RBS controls to the promoter ones.
- Riboswitches do not increase the circuit complexity.

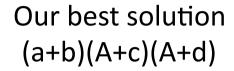
#### Our tool

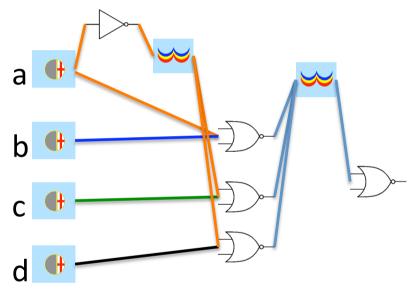
- The truth table is the only input.
- All the schemes compatible with POS and SOP formulas are computed (less than 1s up to 8s).
- They are ranked according to their complexity score.
- The user can choose a solution: this is built by parts, pools, and device composition and encoded in MDL (Model Definition Language) to be visualized in ProMoT (http://www.mpimagdeburg.mpg.de/projects/promot/).



#### Comparison with RNAi-based design

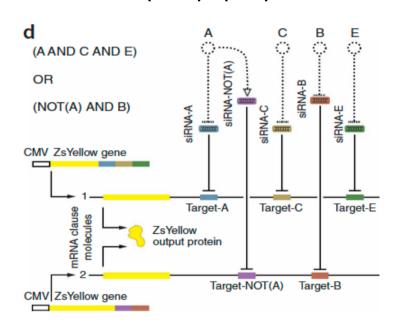
(Rinaudo et al., Nat. Biotech., 25, 795, 2007)





Circuit	Score	Α	R	RNA
Our best POS	2	0	2	0
Our best SOP	4	2	1	1
Rinaudo	5	0	0	5

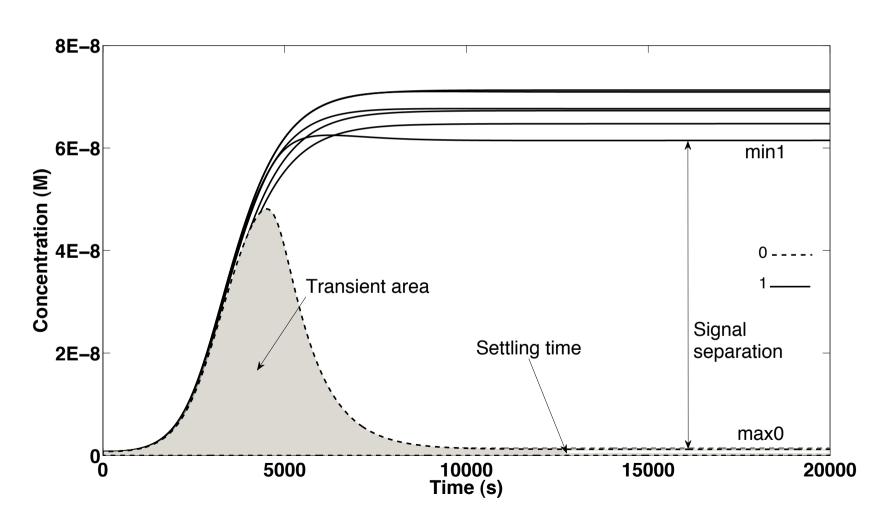
## Rinaudo's solution (acd)+(Ab)



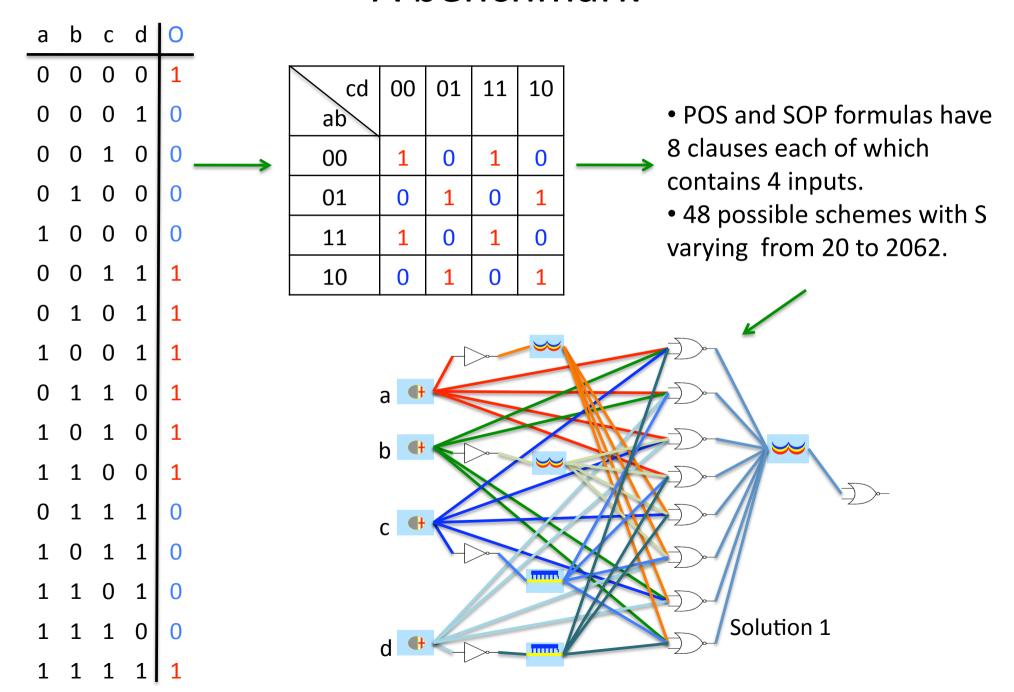
Our tool found 15 designs with complexity lower than 5.

#### How do these circuits work?

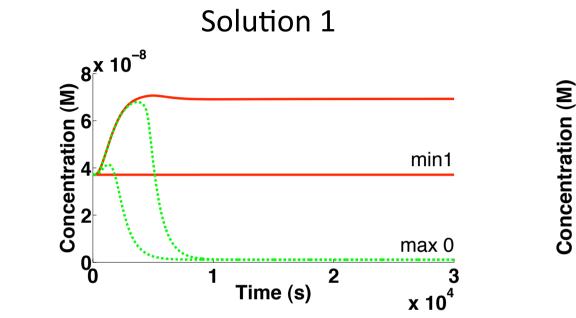
 Circuit performance is estimated through signal separation and transient calculation and depends both on structure and parameter values.

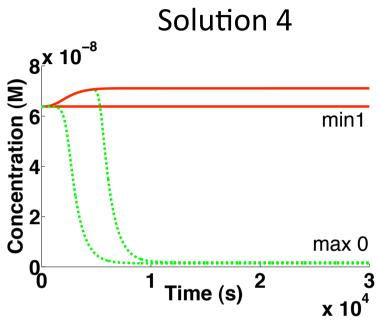


#### A benchmark



#### Comparison of two possible solutions



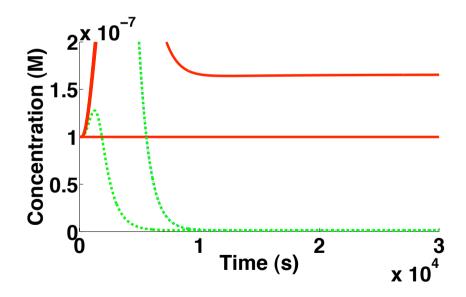


Solution	Rank	Score	Α	R	RNA	Gene	Separation	Transient
1	1	20	2	5	2	17	36 nM	3709
4	25	548	10	6	4	21	62.1 nM	5112

Higher complexity seems to guarantee better performance.

## Improving the performance

- The signal separation is mostly influenced by parameters belonging to the **final gate**.
- Tuning only one parameter (the strength of the promoter in the final gate) the signal separation can be drastically amplified.



 Stochastic algorithms can be avoided but a good set of default parameter values is required.

### Conclusion and future work

- Automatic design of digital synthetic gene circuits via the Karnaugh map method.
- Circuit structure calculation does not require any optimization procedure.
- Theoretical new design of Boolean gates where promoter and RBS are simultaneously.
- Computer simulations show an unequivocal signal separation between 0/1 outputs with our choice of default parameter values.
- Insertion of other translation regulation mechanisms.
- Extension to eukaryotic cells.
- MAIN GOAL: Wet-lab implementation of single gates and more complex circuits.